

# **The Role of Biofilms in Retail Settings**

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## Estimated annual human health burden of selected known foodborne diseases, United States

Pathogen	Illnesses	Deaths	Case-fatality
<i>Campylobacter</i>	1,322,137	119	0.1%
<i>Salmonella</i>	1,229,007	452	0.5%
<i>E. coli</i> O157:H7	96,534	31	0.5%
<i>L. monocytogenes</i>	1662	266	15.9%



Scallan, et al., *Emerging Infectious Diseases*, 2011

# *Listeria monocytogenes* and Retail: A Challenge

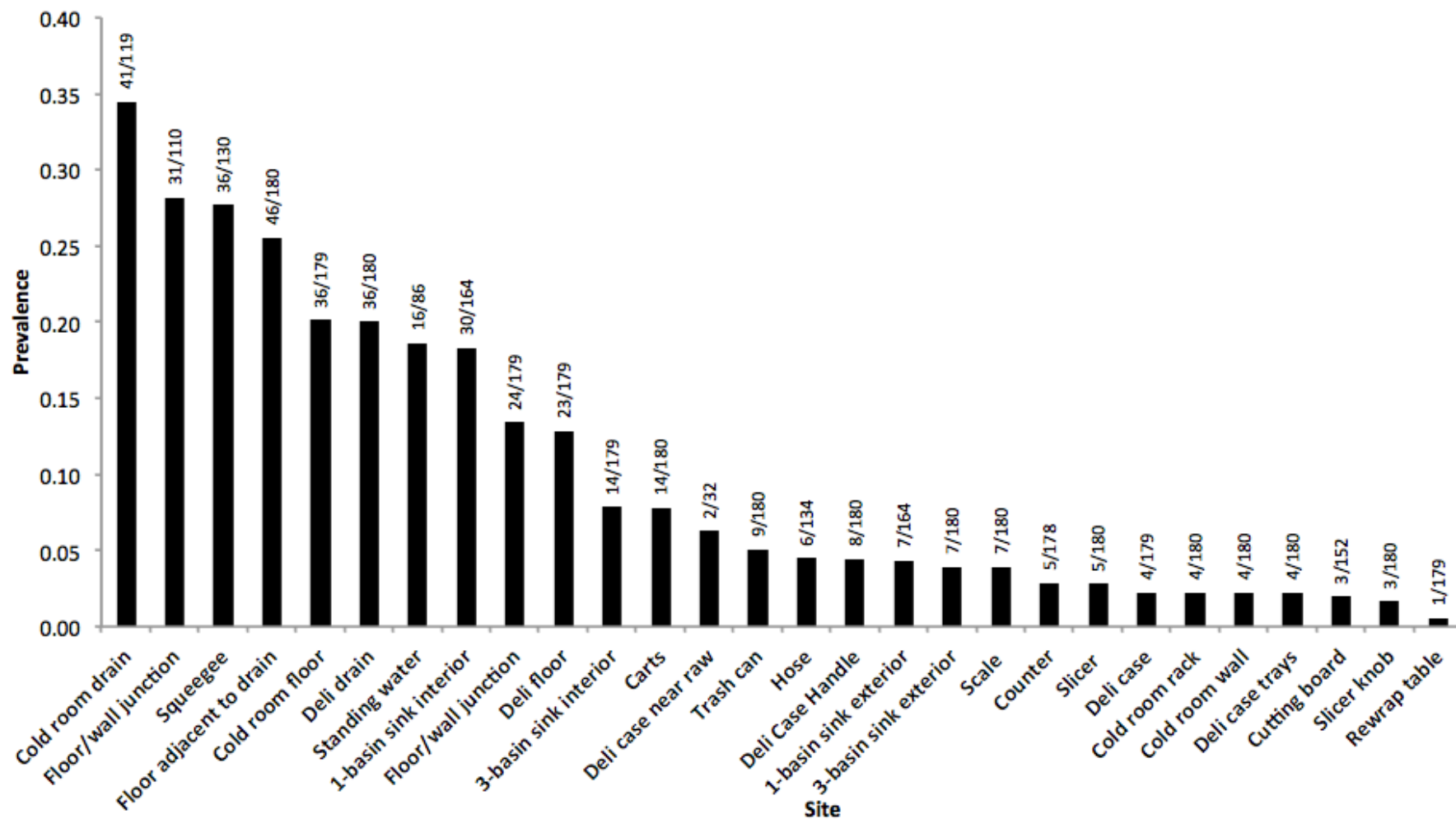
- Ready-to-eat foods: a common source of listeriosis <sup>1</sup>
  - 83% of listeriosis cases from RTE deli meats from deli meats sliced at retail<sup>1</sup>
    - Post-heat treatment contamination<sup>2</sup>
- *L. monocytogenes* prevalence in delis<sup>3</sup>
  - 14.2% of NFCS
  - 4.5% of FCS



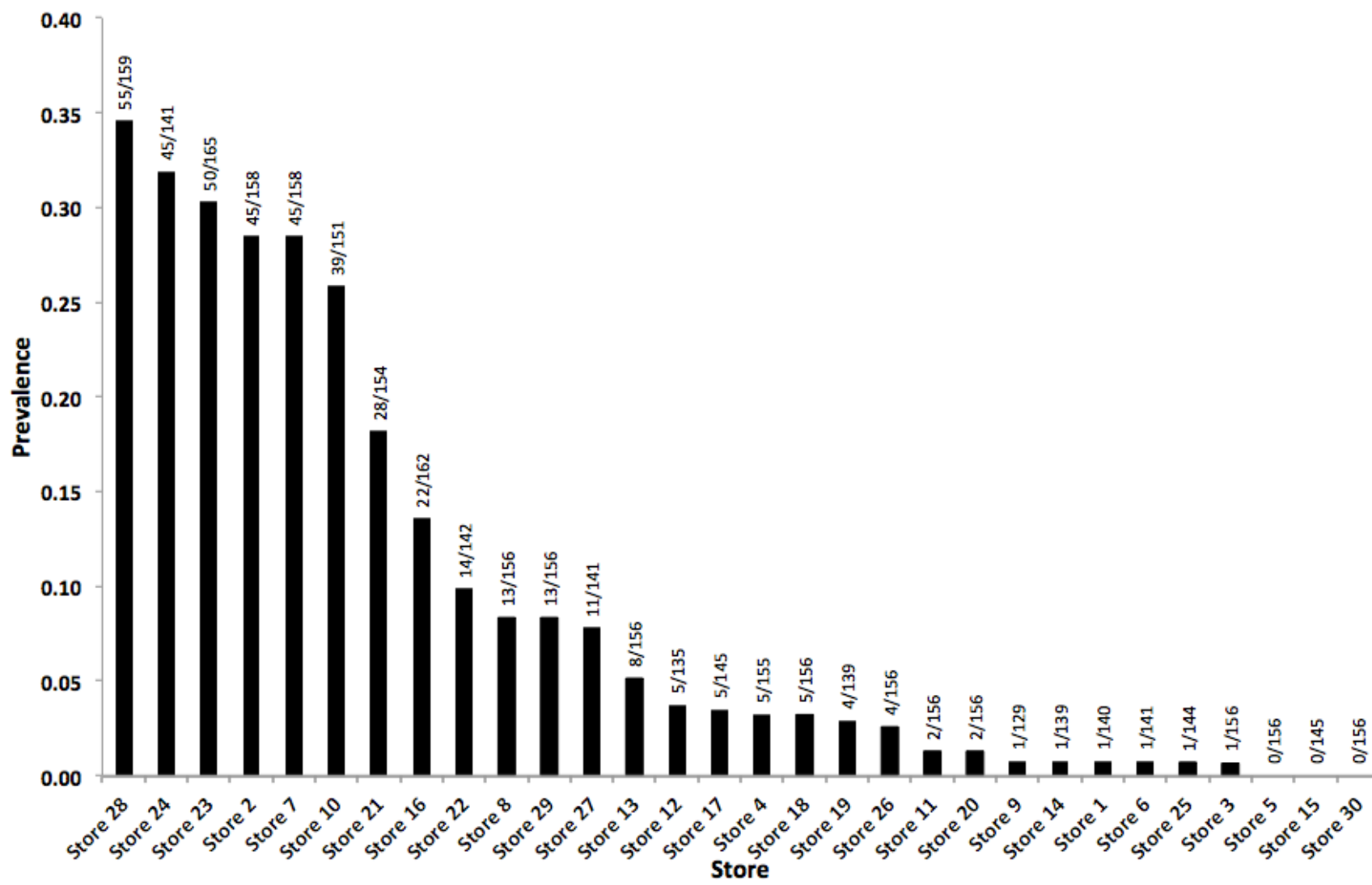
# Persistent and Transient Strains in Delis

- Strain: unique PFGE pulsotype
- Persistent strain: *L. monocytogenes* with PFGE pattern in the same store for  $\geq 3$  separate months
- Transient strain: *L. monocytogenes* with PFGE pattern in the same store for  $< 3$  separate months

# Prevalence of LM by Site



# Prevalence of LM by Store



	April	May	June	July	August	September	October	November	December
<b>Food Contact Sites</b>									
Slicer	-	-	-	-	-	-	-	-	-
Deli case	NT	NT	NT	-	-	-	-	-	-
Deli case near raw meat	NT	NT	NT	-	-	-	-	-	-
Deli case trays	NT	NT	NT	-	-	-	-	-	-
3-basin sink interior	NT	NT	NT	CU-57,267	-	-	-	-	-
1-basin sink interior	NT	NT	NT	CU-258,69	-	-	-	CU-294,321	-
Cold room rack	-	-	-	-	-	-	-	-	-
Cutting board	NT	NT	NT	NT	-	-	NT	NT	-
Rewrap table	NT	NT	NT	-	-	-	-	-	-
Counter	NT	NT	NT	-	-	-	-	-	-
<b>Non-food contact sites</b>									
3-basin sink exterior	NT	NT	NT	-	-	-	-	-	-
Floor/wall junction (3-basin)	CU-258,69	CU-258,69	CU-258,69	CU-258,69	CU-258,69	CU-8,96	LM	CU-258,69	CU-258,69
1-basin sink exterior	NT	NT	NT	CU-258,69	-	-	LM	-	CU-258,69
Floor/wall junction (1-basin)	NT	NT	NT	CU-258,69	-	-	LM	CU-258,69	CU-258,69
Deli drain	NT	NT	NT	CU-258,69	CU-258,333	-	CU-258,69	CU-258,69	CU-258,69
Floor adjacent to drain	-	CU-258,69	CU-258,69	CU-258,69	CU-258,69	-	-	CU-258,69	CU-258,69
Deli floor	NT	NT	NT	CU-258,69	-	-	-	CU-258,69	-
Cold room floor	NT	NT	NT	CU-258,69	CU-295,329	-	CU-258,69	CU-258,69	CU-258,69
Cold room wall	CU-258,69	-	-	-	-	-	-	-	-
Cold room drain	NT	NT	NT	CU-258,69	CU-258,69	-	CU-258,69	CU-258,69	CU-258,69
Standing water	NT	NT	NT	NT	-	-	NT	NT	-
Squeegee	NT	NT	NT	CU-258,69	CU-258,69	-	CU-258,69	CU-258,69	CU-258,69
Cart Wheel	-	-	CU-258,69	CU-258,69	-	-	-	-	-
Hose	NT	NT	NT	-	CU-258,69	-	-	-	-
Trash can	-	-	CU-258,69	-	CU-258,69	-	-	-	-
<b>Transfer Points</b>									
Slicer knob	-	-	-	-	-	-	-	-	-
Case handle	-	-	-	-	-	-	-	-	-
Scale	NT	NT	NT	-	-	-	-	-	-

## Phenotypic Characteristics of LM

- Biofilms
  - Protect LM against environmental stress<sup>4</sup>
  - Form on various food contact surfaces<sup>5</sup>
  - Potential cause of contamination<sup>6</sup>
  - Influenced by: strains, properties of surfaces, temperatures, growth media, and the presence of other microorganisms<sup>7,8</sup>

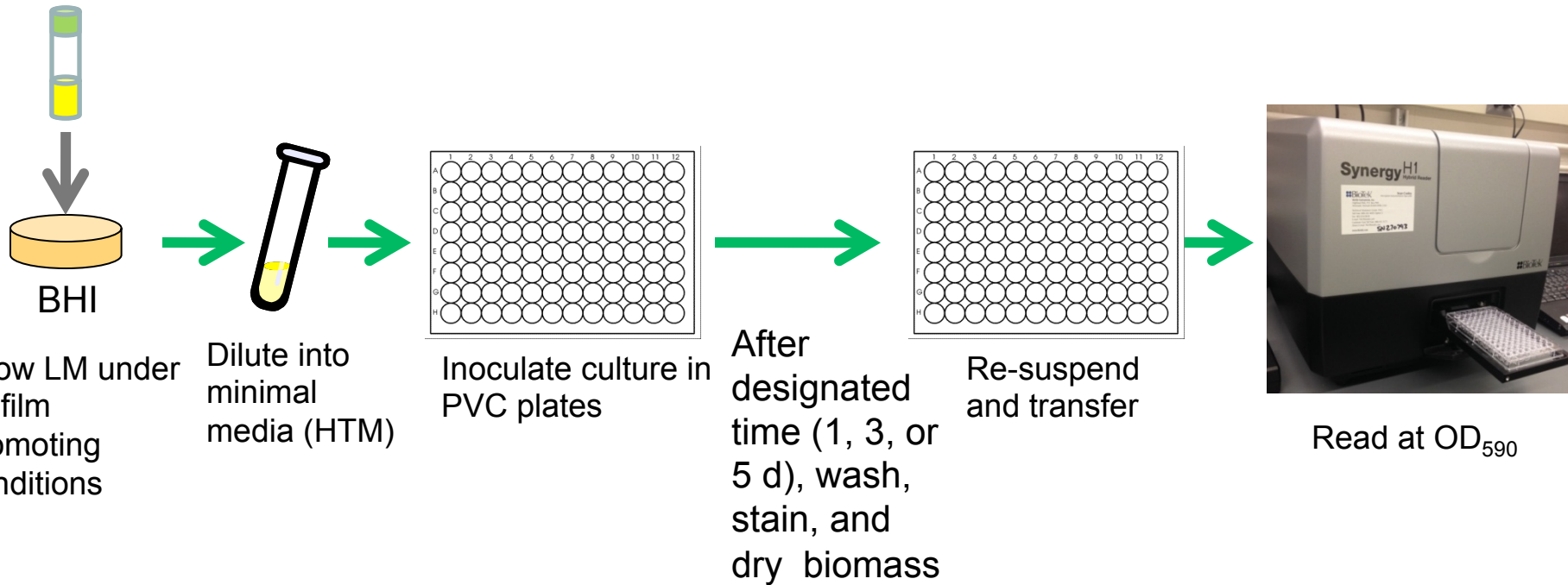


## Phenotypic Characteristics of *L. monocytogenes* Retail Isolates

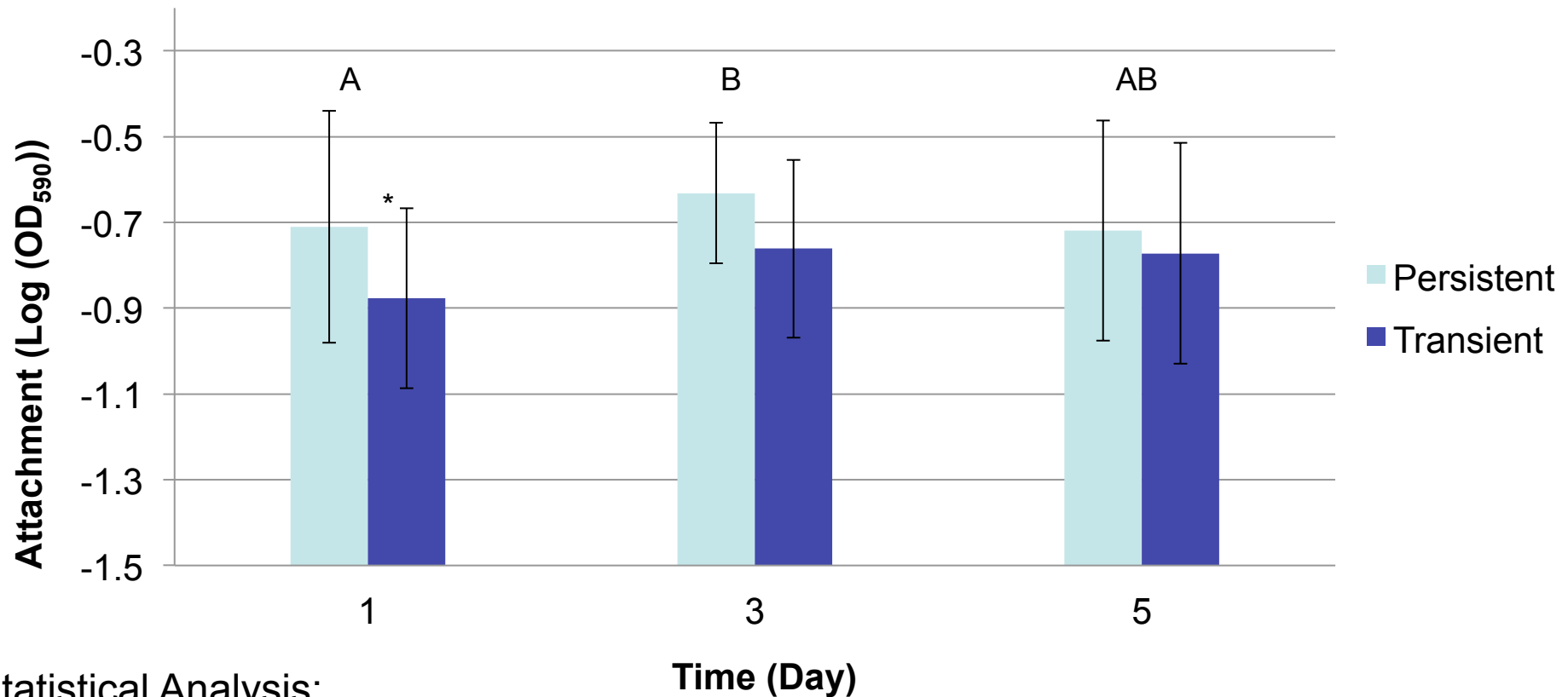
- Hypothesis:
  - *L. monocytogenes* persistent strains are better at forming biofilms
  - There is relationship between biofilm formation and sanitizer tolerance
- Goals of this Work:
  - To assess the ability of 23 persistent strains and 73 transient strains
    - Attachment to abiotic surfaces (indicator of biofilm formation)
    - Sanitizer tolerance
    - Relationship between attachment ability and sanitizer tolerance

# Attachment Assay

Adapted from Lemon et al. 2007, Chen et al. 2013



## Persistent Strains Attach to Abiotic Surfaces Better than Transient Strains

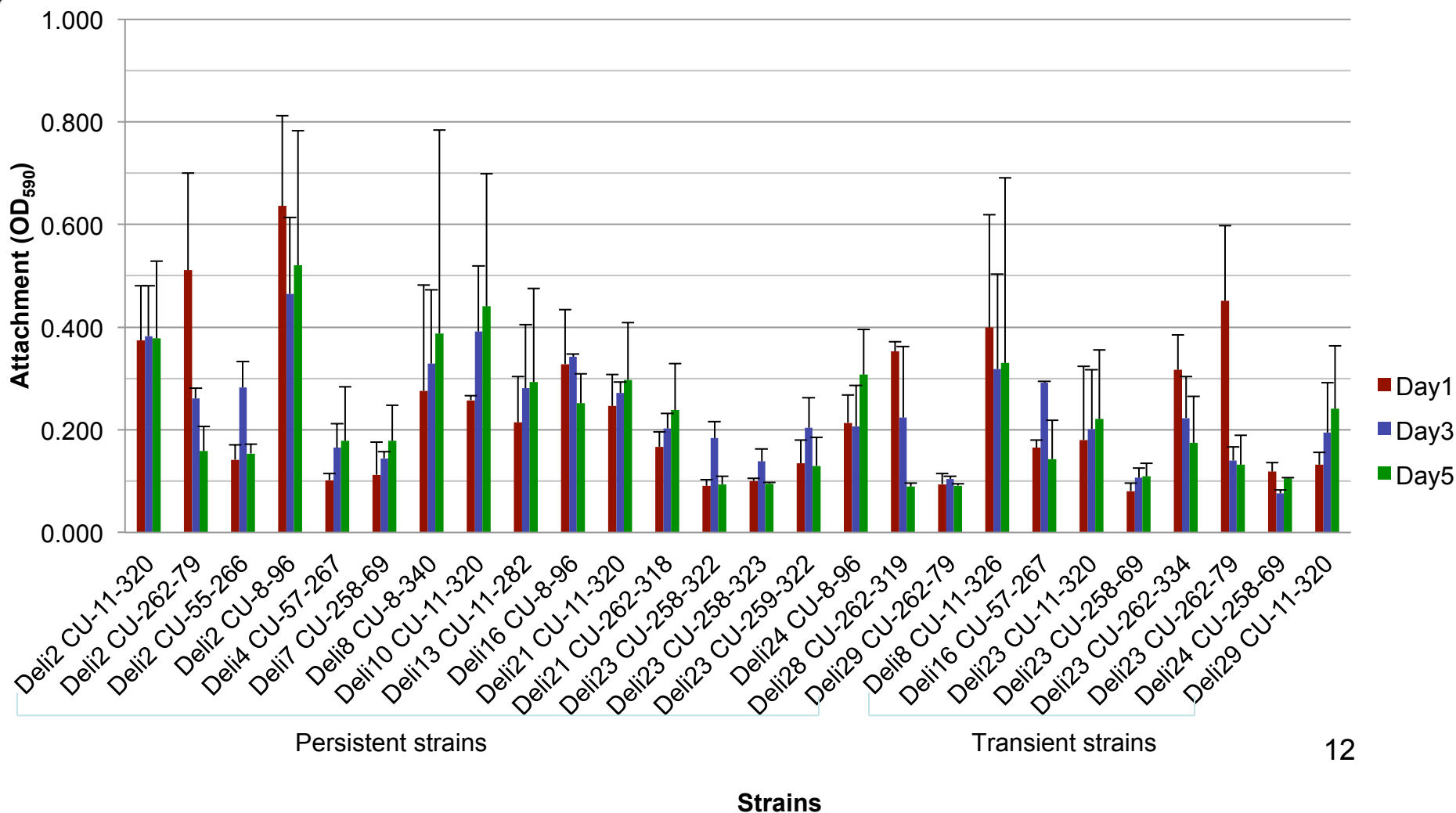


Statistical Analysis:

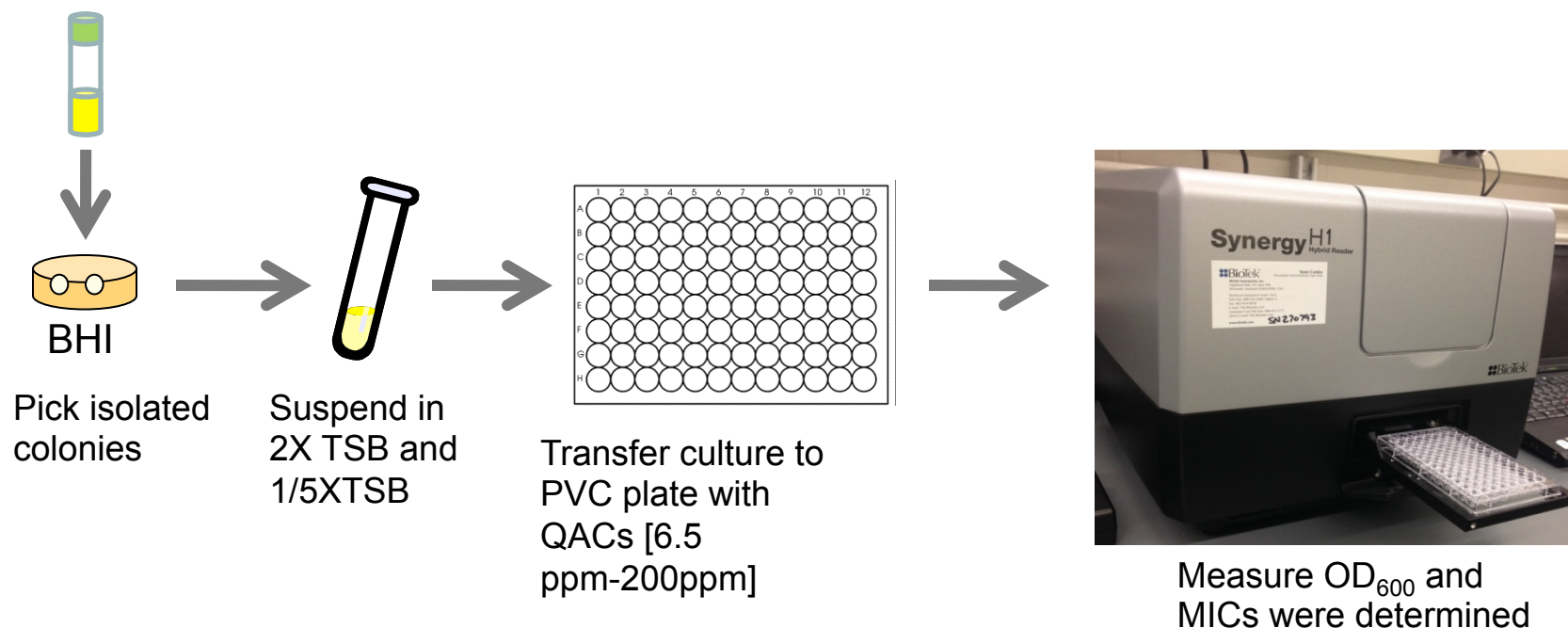
Generalized linear model (GLM):  $\text{Log}(\text{OD}_{590}) = \text{"day", "type", and "day*type"}$   
 LS Means to determine p-value for day\*type

\* Denotes significance at  $P < 0.05$

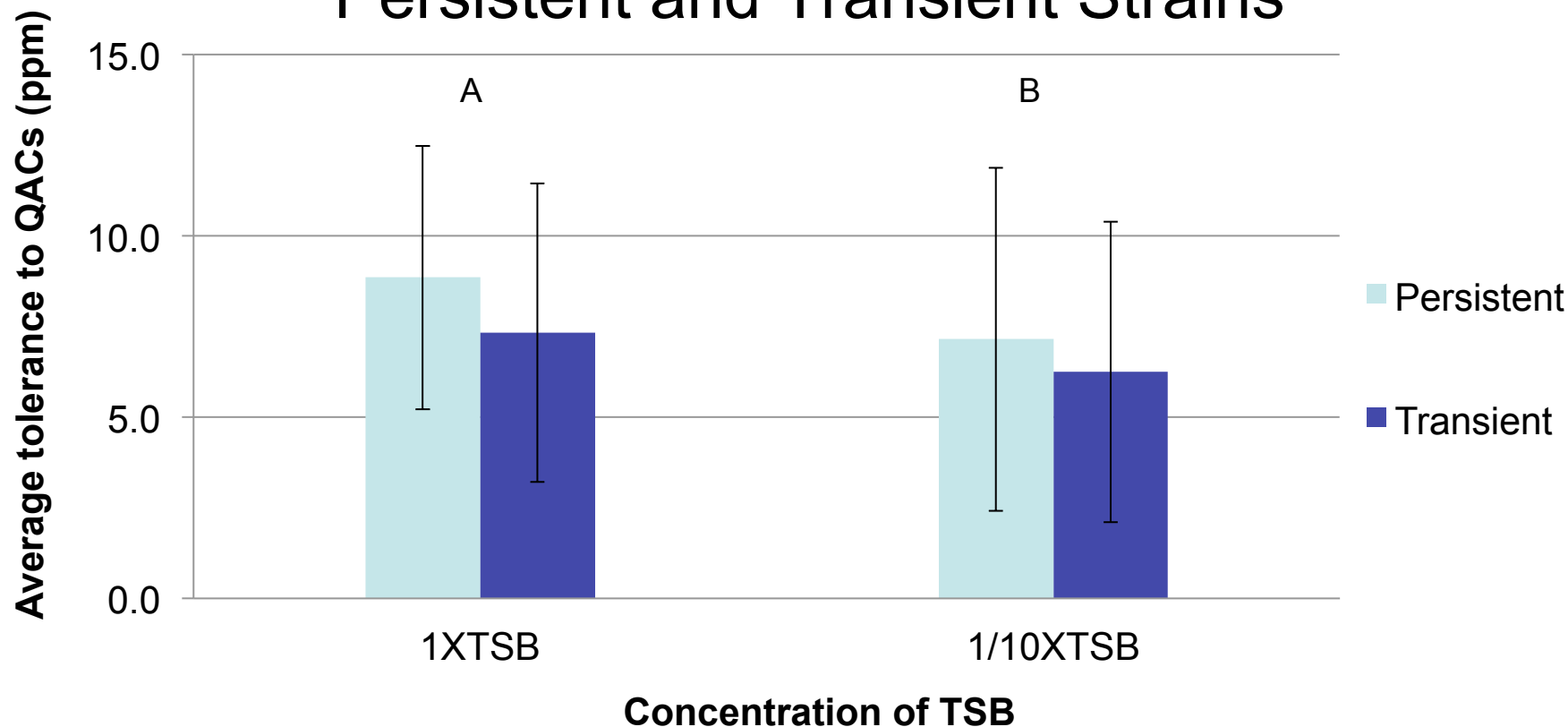
## Isolates Within a Strain Display Significant Variation in Attachment Ability



# Sanitizer Tolerance Assay



## No Significant Difference between Persistent and Transient Strains



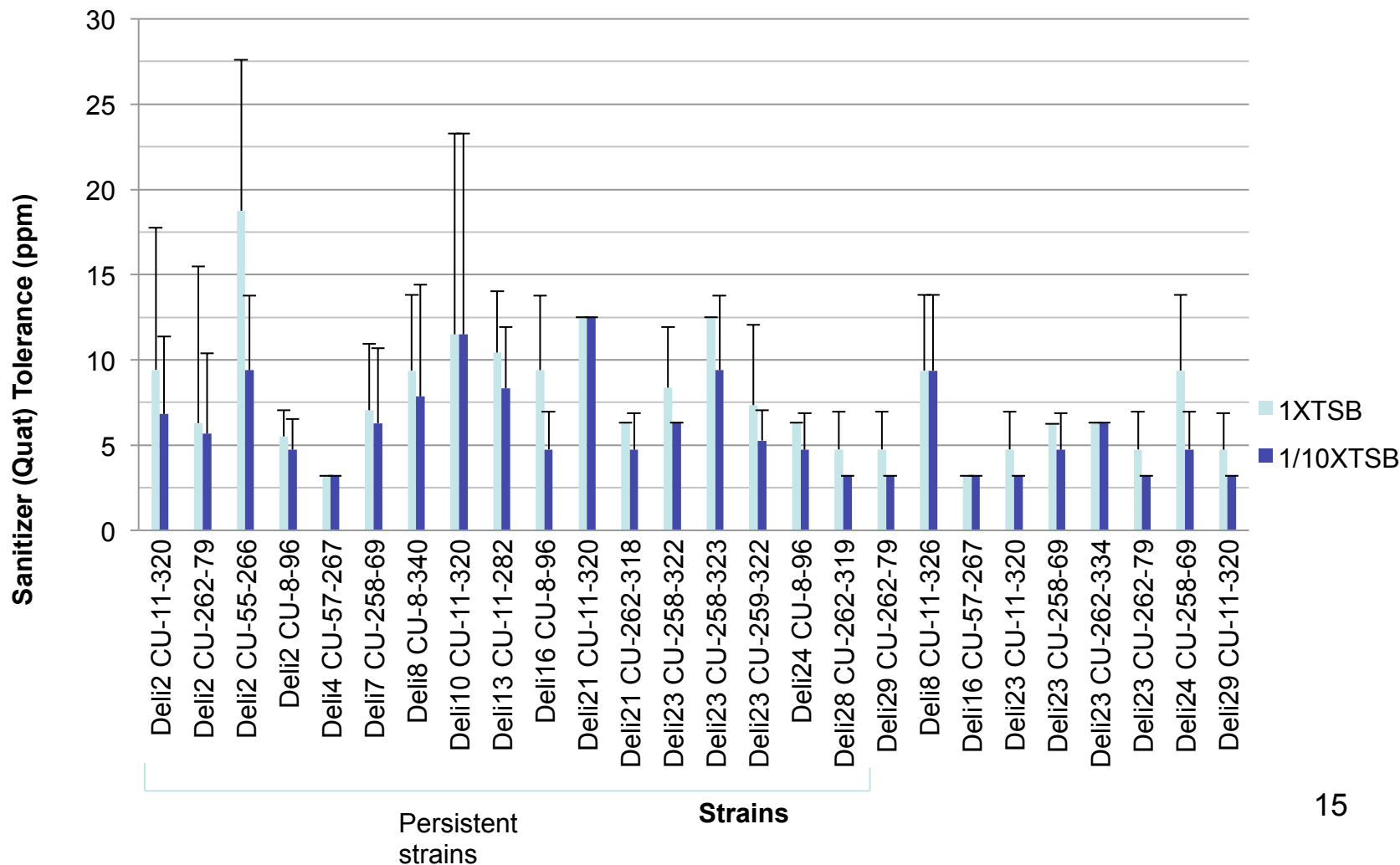
Statistical Analysis:

GLIMMIX: tolerance = "nutrient", "type", and "nutrient\*type"

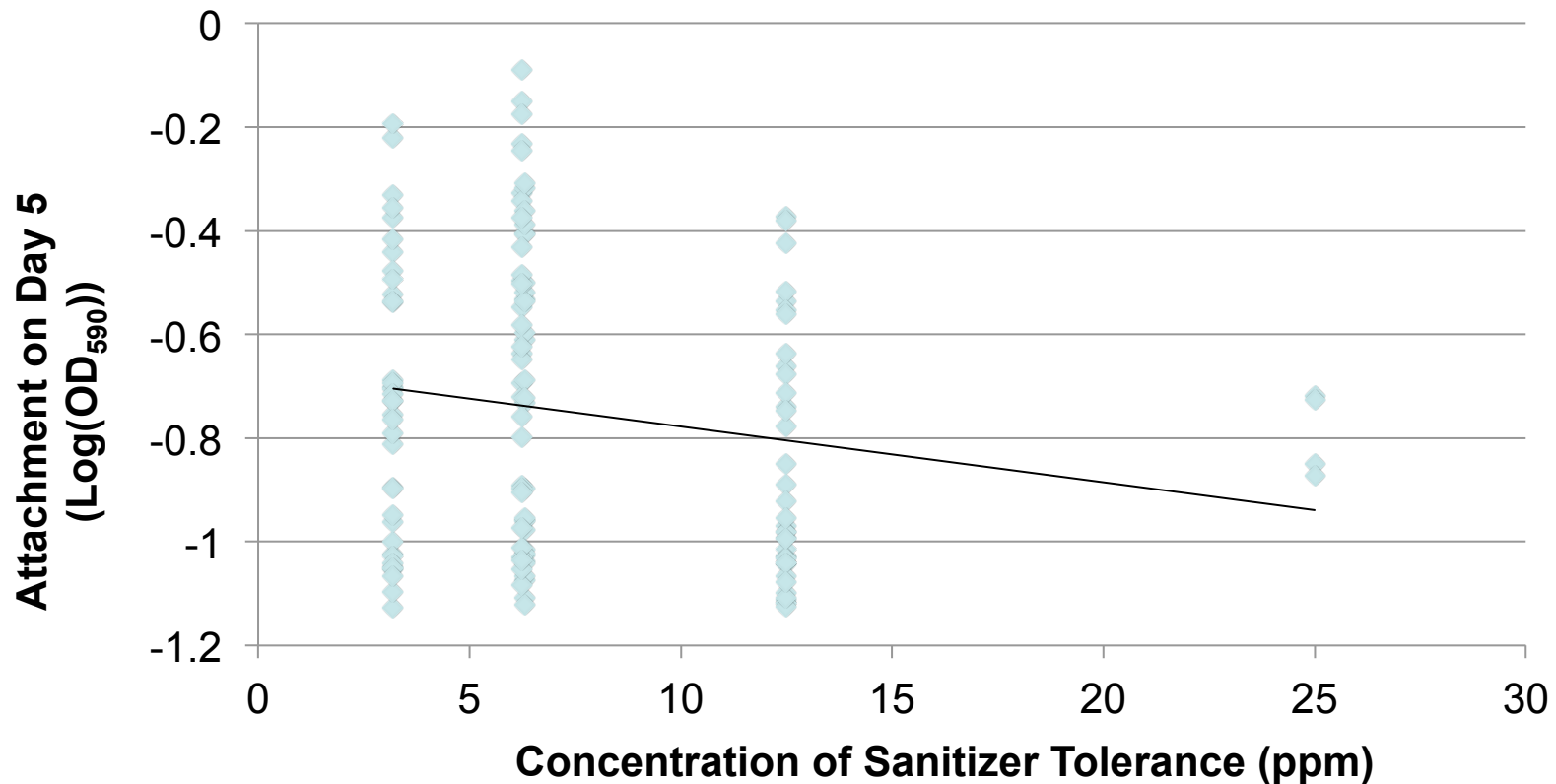
LS Means to determine p-value for "nutrient\*type"

\* Denotes significance at  $P < 0.05$

## Isolates Within a Strain Display Significant Variation in Sanitizer Tolerance



# Attachment Ability is Negatively Correlated with Sanitizer Tolerance after Long-term Incubation in Nutrient Rich Conditions



Statistical Analysis:

Linear regression model:  $\text{Log(OD}_{590}\text{)} \text{ (on different day)} = \text{sanitizer tolerance (under different nutrient conditions)}$

\* Denotes significance at  $P < 0.05$



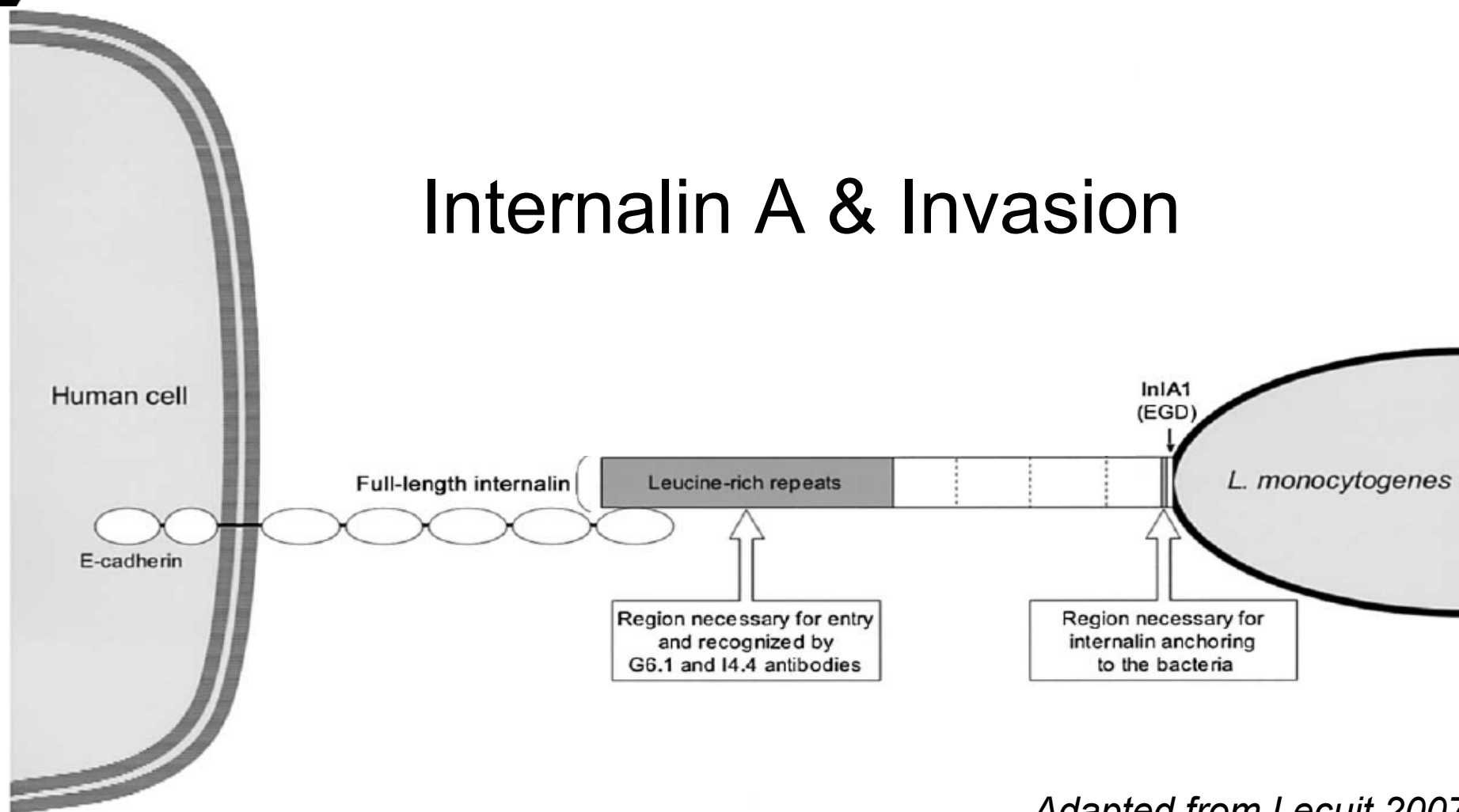
## Conclusion

- Persistent LM strains were better at attaching to abiotic surfaces (PVC) than transient strains
- Persistent strains and transient strains displayed similar levels of sanitizer tolerance
  - LM was more tolerant to QACs under nutrient rich conditions than in nutrient limiting conditions
- A negative correlation was found between enhanced cell attachment on day 5 and sanitizer tolerance under nutrient rich conditions
- Both attachment ability and sanitizer tolerance varied widely among isolates of the same strain

## Key questions:

- What proportion of isolates in the retail deli environment are virulence attenuated?
  - Food contact surfaces vs non-food contact surfaces?
  - Persistence of virulence attenuated mutants compared to WT strains?

# Internalin A & Invasion



## Virulence Potential of *L. monocytogenes* from delis

- Up to 45% of isolates from RTE foods have *inlA* premature stop codons
  - Strongly decreased invasiveness of mutant (<10% of WT) in cell culture
  - Rarely found in isolates from clinical cases



Van Stelten et al., 2010;  
Van Stelten et al., 2008

## PMSC Screen

- 981 isolates (Phases I-V)
  - Test for full length *internalin A* vs PMSC
    - 18 known *inlA* PMSCs
    - PCR-based assay:

Gene amplification



Single nucleotide  
extension PCR



Capillary  
electrophoresis

Van Stelten et al, 2008;  
Van Stelten et al, 2010

## *inlA* PMSCs are Rare Among Deli Isolates

- 15/981 isolates had *inlA* PMSCs
- ~5% of FCS isolates contained PMSCs
  - 8/15 PMSCs associated with FCS isolates
- <1% of NFCS isolates contained PMSCs

## *inlA* PMSCs Not Common in Delis with High *L. monocytogenes* Prevalence

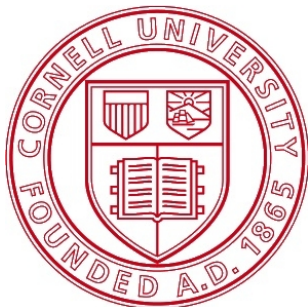
- PMSCs more common in moderate prevalence delis (1-10% average LM+ environmental samples)
  - 11.9% of isolates vs ~2% of isolates from high prevalence stores
  - 8/15 PMSC+ isolates from moderate prevalence delis
- PMSCs significantly associated with FCS isolates from moderate prevalence stores
- **PMSCs always found in transient isolates**

Store Number <sup>a</sup>	LM Prevalence <sup>b</sup>	Total PMSC <sup>d</sup>	FCS PMSC <sup>f</sup>	NFCS PMSC <sup>h</sup>	PFGE Patterns of PMSC+ isolates <sup>i</sup>	PMSC Locations <sup>j</sup>	PMSC Type <sup>k</sup>
Store 2	High	2	0	2	CU-82-215	Floor/wall junction under 1-basin sink (NFCS)	PMSC-4
					CU-341-79	Cold room drain (NFCS)	PMSC-3
Store 16	High	1	0	0	CU-100-140	Deli case handle (TP)	PMSC-4
Store 23	High	3	2	1	CU-296-330	1-basin deli sink interior (FCS)	PMSC-4
					CU-82-215	Standing water (NFCS)	
Store 4	Moderate	1	0	1	CU-80-218	Trash cans (NFCS)	PMSC-4
Store 8	Moderate	1	1	0	CU-200-227	1-basin deli sink interior (FCS)	PMSC-4
Store 11	Moderate	1	1	0	CU-200-227	3-basin deli sink interior (FCS)	PMSC-4
Store 17	Moderate	1	0	1	CU-81-215	Trash cans (NFCS)	PMSC-4
Store 19	Moderate	1	1	0	CU-100-140	Countertop (FCS)	PMSC-3
Store 20	Moderate	1	1	0	CU-200-227	3-basin deli sink interior (FCS)	PMSC-4
Store 22	Moderate	2	1	1	CU-180-231	Deli case (FCS)	PMSC-4
					CU-258-332	Deli floor (NFCS)	
Store 13	Low	1	1	0	CU-200-227	Cold room racks (FCS)	PMSC-4
<b>Total:</b>		<b>11</b>	<b>7</b>	<b>4</b>			



## Persistent Isolates Are More Likely to Form Biofilms and Less Likely to Have *inlA* PMSCs

- Persistent *L. monocytogenes* strains may be better adapted for survival in retail delis through biofilm formation
- Biofilm-forming isolates are less likely to have virulence-attenuating mutations
  - Risk for cross-contamination of surfaces



**United States Department of Agriculture**  
Food Safety and Inspection Service



# References

1. FSIS, FDA-FSIS Quantitative Risk Assessment for *Listeria monocytogenes* in Ready-to-Eat Foods. 2003.
2. Tompkin, R. Bruce, et al., Guidelines to prevent post-processing contamination from *Listeria monocytogenes*. Dairy Food and Environmental sanitation 19 (1999): 551-603.
3. Simmons, C., M. Stasiewicz, S. Roof, S. Hammons, E. Wright, S. Worchoki, M. Wiedmann, and H. and Oliver. unpublished. Prevalence and Persistence of *L. monocytogenes* in Retail Delis in Three States. *In preparation for submission to J Food Prot.*
4. Djordjevic, D., M. Wiedmann, and L. McLandsborough. 2002. Microtiter plate assay for assessment of *Listeria monocytogenes* biofilm formation. *Applied and Environmental Microbiology*. 68:2950-2958.
5. Truelstrup Hansen, L., and B. F. Vogel. 2011. Desiccation of adhering and biofilm *Listeria monocytogenes* on stainless steel: Survival and transfer to salmon products. *International Journal of Food Microbiology*. 146:88-93.
6. Tompkin, R. B. 2002. Control of *Listeria monocytogenes* in the food-processing environment. *Journal of Food Protection®*. 65:709-725.
7. Palmer, J., S. Flint, and J. Brooks. 2007. Bacterial cell attachment, the beginning of a biofilm. *Journal of industrial microbiology & biotechnology*. 34:577-588.
8. Pan, Y., F. Breidt, and S. Kathariou. 2006. Resistance of *Listeria monocytogenes* biofilms to sanitizing agents in a simulated food processing environment. *Applied and Environmental Microbiology*. 72:7711-7717.
9. Fraise, A., J.-Y. Maillard, and S. Sattar. 2012. Russell, Hugo and Ayliffe's Principles and Practice of Disinfection, Preservation and Sterilization. John Wiley & Sons.
10. Romanova, N., P. Wolffs, L. Brovko, and M. Griffiths. 2006. Role of efflux pumps in adaptation and resistance of *Listeria monocytogenes* to benzalkonium chloride. *Applied and environmental microbiology*. 72:3498-3503.
11. Tezel, U., J. A. Pierson, and S. G. Pavlostathis. 2006. Fate and effect of quaternary ammonium compounds on a mixed methanogenic culture. *Water research*. 40:3660-3668.
12. Lunden, J.M., Miettinen M.K., Autio T.J. and Korkeala H.J. 2000. Persistent *Listeria monocytogenes* strains show enhanced adherence to food contact surface after short contact times. *Journal of Food Protection*. 63(9): p. 1204-1207
13. Borucki, M.K, Peppin J.D., White D., Loge F. and Call D.R. 2003. Variation in biofilm formation among strains of *Listeria monocytogenes*. *Applied and Environmental Microbiology*. 69(12): p. 7336-7342

# References

- Endrikat, S., D. Gallagher, et al. (2010). "A comparative risk assessment for *Listeria monocytogenes* in prepackaged versus retail-sliced deli meat." *Journal of Food Protection*® 73(4): 612-619.
- Gokulan, K., S. Khare, et al. (2013). "Impact of plasmids, including those encoding VirB4/D4 type IV secretion systems, on *Salmonella enterica* serovar Heidelberg virulence in macrophages and epithelial cells." *PLoS One* 8(10): e77866.
  - Hammons, S. R., Wang, J., Ray, A.J., Oliver H.F. (unpublished). "Evaluation of deep clean SSOP as a *Listeria monocytogenes* control strategy in retail delis."
  - Nightingale, K. K., R. A. Ivy, et al. (2008). "inlA premature stop codons are common among *Listeria monocytogenes* isolates from foods and yield virulence-attenuated strains that confer protection against fully virulent strains." *Appl Environ Microbiol* 74(21): 6570-6583.
  - Nightingale, K. K., K. Windham, et al. (2005). "Select *Listeria monocytogenes* subtypes commonly found in foods carry distinct nonsense mutations in inlA, leading to expression of truncated and secreted internalin A, and are associated with a reduced invasion phenotype for human intestinal epithelial cells." *Appl Environ Microbiol* 71(12): 8764-8772.
  - Scallan, E., R. M. Hoekstra, et al. (2011). "Foodborne illness acquired in the United States--major pathogens." *Emerg Infect Dis* 17(1): 7-15.
  - Simmons, C., M. J. Stasiewicz, et al. (2014). "*Listeria monocytogenes* and *Listeria* spp. contamination patterns in retail delicatessen establishments in three U.S. states." *J Food Prot* 77(11): 1929-1939.
  - Van Stelten, A. and K. K. Nightingale (2008). "Development and implementation of a multiplex single-nucleotide polymorphism genotyping assay for detection of virulence-attenuating mutations in the *Listeria monocytogenes* virulence-associated gene inlA." *Appl Environ Microbiol* 74(23): 7365-7375.
  - Van Stelten, A., J. M. Simpson, et al. (2010). "Revelation by single-nucleotide polymorphism genotyping that mutations leading to a premature stop codon in inlA are common among *Listeria monocytogenes* isolates from ready-to-eat foods but not human listeriosis cases." *Appl Environ Microbiol* 76(9): 2783-2790.